

FILE 'MEDLINE, EMBASE, BIOSIS' ENTERED AT 12:58:08 ON 22 AUG 2005

L1 12777 S MENDEZ?/AU OR FINER?/AU  
L2 10971 S (SHUTTLE OR HYBRID) (S) (VECTOR OR PLASMID OR CONSTRUCT)  
L3 463478 S MARKER OR SELECTABLE  
L4 15458 S ADENOVRIUS OR AAV OR "ADENO ASSOCIATED" OR POX OR PAPOVA OR  
L5 20758 S ATTT OR TN7 OR FLP OR LOX OR CRE OR CIRCULARIZATION  
L6 7 S L1 AND L2  
L7 3 DUP REM L6 (4 DUPLICATES REMOVED)  
L8 2 S L7 NOT PY>=2000  
L9 1 S L2 AND L3 AND L4  
L10 1033 S L5 AND L3  
L11 15 S L10 AND L2  
L12 2 S L11 AND YEAST  
L13 7 DUP REM L11 (8 DUPLICATES REMOVED)  
L14 1 DUP REM L12 (1 DUPLICATE REMOVED)  
L15 4 S L13 NOT PY>=2000

L9 ANSWER 1 OF 1 BIOSIS COPYRIGHT (c) 2005 The Thomson Corporation on STN  
ACCESSION NUMBER: 2004:289891 BIOSIS  
DOCUMENT NUMBER: PREV200400288085  
TITLE: Transfection-free and scalable recombinant **AAV**  
vector production using HSV/**AAV** hybrids.  
AUTHOR(S): Booth, M. J.; Mistry, A.; Li, X.; Thrasher, A.; Coffin, R.  
S. [Reprint Author]  
CORPORATE SOURCE: Windeyer InstDept Immunol and Mol Pathol, Univ Coll London,  
46 Cleveland St, London, W1T 4JF, England  
SOURCE: Gene Therapy, (May 2004) Vol. 11, No. 10, pp. 829-837.  
print.  
ISSN: 0969-7128 (ISSN print).  
DOCUMENT TYPE: Article  
LANGUAGE: English  
ENTRY DATE: Entered STN: 16 Jun 2004  
Last Updated on STN: 16 Jun 2004

AB **Adeno-associated virus (AAV)** vectors are highly efficient tools for use in gene therapy. Current production methods rely on plasmid transfection and are not generally considered amenable to scale-up. To improve recombinant **AAV** (rAAV) vector production in terms of both final titre and simplicity, we constructed recombinant herpes simplex virus (HSV) vectors, either disabled (ICP27 deleted) or nondisabled, encoding the **AAV** rep and cap genes. We also integrated an rAAVGFP construct into the nondisabled vector and also into a second pair of HSV vectors ( disabled and nondisabled) not expressing rep and cap. Transgene incorporation and expression was confirmed by Southern and Western blot, respectively. Optimal double-infection ratios were established for disabled and nondisabled pairs of rep/cap-expressing and rAAVGFP-containing vectors, resulting in up to  $1.55 \times 10^{12}$  rAAV capsids and  $4 \times 10^8$  expression units from approximately  $1 \times 10^7$  BHK producer cells. Functionality of the prepared vector was confirmed by the detection of abundant green fluorescent protein (GFP) expression following injections of rAAV preparations into the rat brain. This paper therefore describes a simple, efficient, and transfection-free rAAV production process based on the use of HSV and not relying on a proviral cell line that, with appropriate scale-up, could yield quantities of rAAV sufficient for routine clinical use.

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L14 ANSWER 1 OF 1 MEDLINE on STN DUPLICATE 1  
ACCESSION NUMBER: 88210531 MEDLINE  
DOCUMENT NUMBER: PubMed ID: 3449223  
TITLE: Terminal segment of Kluyveromyces lactis linear DNA  
**plasmid** pGKL2 supports autonomous replication of  
**hybrid** plasmids in Saccharomyces cerevisiae.  
AUTHOR: Fujimura H; Hishinuma F; Gunge N  
CORPORATE SOURCE: Mitsubishi Kasei Institute of Life Sciences, Tokyo, Japan.  
SOURCE: Current genetics, (1987) 12 (2) 99-104.  
Journal code: 8004904. ISSN: 0172-8083.  
PUB. COUNTRY: United States  
DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
LANGUAGE: English  
FILE SEGMENT: Priority Journals  
OTHER SOURCE: GENBANK-X05601  
ENTRY MONTH: 198806  
ENTRY DATE: Entered STN: 19900308  
Last Updated on STN: 19990129  
Entered Medline: 19880617

AB By use of linear DNA **plasmid** pGKL2 from the **yeast**  
Kluyveromyces lactis we have constructed **hybrid** plasmids  
carrying a LEU2 gene of Saccharomyces cerevisiae as a **selectable**  
**marker**. The replication properties of hybrid plasmids in yeasts  
were investigated. We demonstrated that the insertion of a LEU2 gene into  
pGKL2 resulted in **circularization** of the hybrid plasmids and  
pGKL2 segment supported autonomous replication of the plasmids. Moreover,  
the **hybrid** plasmids propagated autonomously, independently of  
the presence of the natural pGKL2 **plasmid**.

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L15 ANSWER 1 OF 4 MEDLINE on STN  
 ACCESSION NUMBER: 1999200479 MEDLINE  
 DOCUMENT NUMBER: PubMed ID: 10102356  
 TITLE: pBECKS2000: a novel plasmid series for the facile creation of complex binary vectors, which incorporates "clean-gene" facilities.  
 AUTHOR: McCormac A C; Elliott M C; Chen D F  
 CORPORATE SOURCE: The Norman Borlaug Institute for Plant Science Research, De Montfort University, Scraptoft, Leicester, UK.. amccorma@dmu.ac.uk  
 SOURCE: Molecular & general genetics : MGG, (1999 Mar) 261 (2) 226-35.  
 Journal code: 0125036. ISSN: 0026-8925.  
 PUB. COUNTRY: GERMANY: Germany, Federal Republic of  
 DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
 LANGUAGE: English  
 FILE SEGMENT: Priority Journals  
 ENTRY MONTH: 199904  
 ENTRY DATE: Entered STN: 19990511  
 Last Updated on STN: 19990511  
 Entered Medline: 19990426

AB A new plasmid series has been created for Agrobacterium-mediated plant transformation. The pBECKS2000 series of binary vectors exploits the **Cre**/ loxP site-specific recombinase system to facilitate the construction of complex T-DNA vectors. The new plasmids enable the rapid generation of T-DNA vectors in which multiple genes are linked, without relying on the availability of purpose-built cassette systems or demanding complex, and therefore inefficient, ligation reactions. The vectors incorporate facilities for the removal of transformation markers from transgenic plants, while still permitting simple in vitro manipulations of the T-DNA vectors. A '**shuttle**' or intermediate **plasmid** approach has been employed. This permits independent ligation strategies to be used for two gene sets. The intermediate plasmid sequence is incorporated into the binary vector through a plasmid co-integration reaction which is mediated by the **Cre**/loxP site-specific recombinase system. This reaction is carried out within Agrobacterium cells. Recombinant clones, carrying the co-integrative binary plasmid form, are selected directly using the antibiotic resistance **marker** carried on the intermediate plasmid. This strategy facilitates production of co-integrative T-DNA binary vector forms which are appropriate for either (1) transfer to and integration within the plant genome of target and **marker** genes as a single T-DNA unit; (2) transfer and integration of target and **marker** genes as a single T-DNA unit but with a **Cre**/loxP facility for site-specific excision of **marker** genes from the plant genome; or (3) co-transfer of target and **marker** genes as two independent T-DNAs within a single-strain Agrobacterium system, providing the potential for segregational loss of **marker** genes.

L15 ANSWER 2 OF 4 MEDLINE on STN  
 ACCESSION NUMBER: 93323197 MEDLINE  
 DOCUMENT NUMBER: PubMed ID: 8392598  
 TITLE: Efficient generation of infectious recombinant baculoviruses by site-specific transposon-mediated insertion of foreign genes into a baculovirus genome propagated in Escherichia coli.  
 AUTHOR: Luckow V A; Lee S C; Barry G F; Olins P O  
 CORPORATE SOURCE: Cellular and Molecular Biochemistry, Monsanto Corporate Research, Chesterfield, Missouri 63198.  
 SOURCE: Journal of virology, (1993 Aug) 67 (8) 4566-79..  
 Journal code: 0113724. ISSN: 0022-538X.  
 PUB. COUNTRY: United States  
 DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
 LANGUAGE: English  
 FILE SEGMENT: Priority Journals  
 ENTRY MONTH: 199308  
 ENTRY DATE: Entered STN: 19930826

Last Updated on STN: 19990129

Entered Medline: 19930816

AB The construction and purification of recombinant baculovirus vectors for the expression of foreign genes in insect cells by standard transfection and plaque assay methods can take as long as 4 to 6 weeks. This period can be reduced to several days by using a novel baculovirus **shuttle vector** (bacmid) that can replicate in *Escherichia coli* as a **plasmid** and can infect susceptible lepidopteran insect cells. The bacmid is a recombinant virus that contains a mini-F replicon, a kanamycin resistance **marker**, and attTn7, the target site for the bacterial transposon Tn7. Expression cassettes comprising a baculovirus promoter driving expression of a foreign gene that is flanked by the left and right ends of Tn7 can transpose to the target bacmid in *E. coli* when Tn7 transposition functions are provided in trans by a helper plasmid. The foreign gene is expressed when the resulting composite bacmid is introduced into insect cells.

L15 ANSWER 3 OF 4 MEDLINE on STN  
ACCESSION NUMBER: 88210531 MEDLINE  
DOCUMENT NUMBER: PubMed ID: 3449223  
TITLE: Terminal segment of *Kluyveromyces lactis* linear DNA **plasmid** pGKL2 supports autonomous replication of **hybrid** plasmids in *Saccharomyces cerevisiae*.  
AUTHOR: Fujimura H; Hishinuma F; Gunge N  
CORPORATE SOURCE: Mitsubishi Kasei Institute of Life Sciences, Tokyo, Japan.  
SOURCE: Current genetics, (1987) 12 (2) 99-104.  
Journal code: 8004904. ISSN: 0172-8083.  
PUB. COUNTRY: United States  
DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
LANGUAGE: English  
FILE SEGMENT: Priority Journals  
OTHER SOURCE: GENBANK-X05601  
ENTRY MONTH: 198806  
ENTRY DATE: Entered STN: 19900308  
Last Updated on STN: 19990129  
Entered Medline: 19880617

AB By use of linear DNA **plasmid** pGKL2 from the yeast *Kluyveromyces lactis* we have constructed **hybrid** plasmids carrying a LEU2 gene of *Saccharomyces cerevisiae* as a **selectable marker**. The replication properties of hybrid plasmids in yeasts were investigated. We demonstrated that the insertion of a LEU2 gene into pGKL2 resulted in **circularization** of the hybrid plasmids and pGKL2 segment supported autonomous replication of the plasmids. Moreover, the **hybrid** plasmids propagated autonomously, independently of the presence of the natural pGKL2 **plasmid**.

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on STN  
ACCESSION NUMBER: 89010830 EMBASE  
DOCUMENT NUMBER: 1989010830  
TITLE: A broad-host-range shuttle system for gene insertion into the chromosomes of Gram-negative bacteria.  
AUTHOR: Barry G.F.  
CORPORATE SOURCE: Plant Microbiology Group, Biological Sciences Department, Monsanto Company, St. Louis, MO 63198, United States  
SOURCE: Gene, (1988) Vol. 71, No. 1, pp. 75-84.  
ISSN: 0378-1119 CODEN: GENED6  
COUNTRY: Netherlands  
DOCUMENT TYPE: Journal  
FILE SEGMENT: 022 Human Genetics  
LANGUAGE: English  
SUMMARY LANGUAGE: English  
ENTRY DATE: Entered STN: 911212  
Last Updated on STN: 911212

AB A deletion derivative of transposon Tn7 containing the *Escherichia coli* lacZY genes as a **selectable marker** for insertion of foreign DNA into the chromosomes of soil bacteria was

improved to facilitate the cloning of additional genes and their insertion by this element. This report describes a series of plasmid vectors that enable this cloning to be carried out in small, high-copy, narrow host-range plasmids. The final Tn element can then be easily moved (by transposition) without further use of restriction enzymes, to plasmids suitable for delivering it to the bacterial chromosome. The very high specificity for insertion of **Tn7** into single locations in bacterial chromosomes has been exploited in the construction of a shuttle system for delivering these **Tn7** elements.

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	Document ID	Title
1	US 20050142562 A1	High throughput generation and screening of fully human antibody repertoire in yeast
2	US 20050124010 A1	Whole cell engineering by mutagenizing a substantial portion of a starting genome combining mutations and optionally repeating
3	US 20050123996 A1	Assembly and screening of highly complex and fully human antibody repertoire in yeast
4	US 20050112141 A1	Compositions and methods for treatment of neoplastic disease
5	US 20050059109 A1	Methods and compositions for polypeptide engineering
6	US 20040245317 A1	Artificial chromosomes that can shuttle between bacteria yeast and mammalian cells
7	US 20040219516 A1	Viral vectors containing recombination sites
8	US 20040214783 A1	Compositions and methods for treatment of neoplastic disease
9	US 20040214277 A1	Methods and compositions for polypeptide engineering
10	US 20040067532 A1	High throughput generation and affinity maturation of humanized antibody

	Document ID	Title
11	US 20040005591 A1	Cloning system for construction of recombinant expression vectors
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13	US 20030186356 A1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
14	US 20030165990 A1	Generation of highly diverse library of expression vectors via homologous recombination in yeast
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16	US 20030027213 A1	Haploid yeast cells transformed with a library of expression vectors encoding a fully human antibody repertoire
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18	US 20020177551 A1	Compositions and methods for treatment of neoplastic disease
19	US 20020051976 A1	METHODS AND COMPOSITIONS FOR POLYPEPTIDE ENGINEERING
20	US 6610472 B1	Assembly and screening of highly complex and fully human antibody repertoire in yeast



	Document ID	Title
21	US 6573098 B1	Nucleic acid libraries
22	US 6551828 B1	Compositions and methods for generating expression vectors through site-specific recombination
23	US 6518065 B1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
24	US 6506603 B1	Shuffling polynucleotides by incomplete extension
25	US 6506602 B1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
26	US 6413774 B1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
27	US 6410271 B1	Generation of highly diverse library of expression vectors via homologous recombination in yeast
28	US 6410246 B1	Highly diverse library of yeast expression vectors
29	US 6406863 B1	High throughput generation and screening of fully human antibody repertoire in yeast

	Document ID	Title
30	US 6395547 B1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
31	US 6372497 B1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
32	US 6344356 B1	Methods for recombining nucleic acids
33	US 6323030 B1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
34	US 6291242 B1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
35	US 6180406 B1	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
36	US 6165793 A	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination
37	US 6117679 A	Methods for generating polynucleotides having desired characteristics by iterative selection and recombination



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